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**Reply to "Modulatory Effects of a Subinhibitory Concentration of  
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aureus* Strains of Sequence Type 30"**

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# Reply to “Modulatory Effects of a Subinhibitory Concentration of Clindamycin in Community-Acquired Methicillin-Resistant *Staphylococcus aureus* Strains of Sequence Type 30”

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**W**e thank Wong and colleagues (1) for taking our work (2) a step further by investigating the impact of a subinhibitory concentration of clindamycin on ST30 strains of community-acquired *Staphylococcus aureus* (CA-MRSA). In our study, we observed that subinhibitory concentrations of the protein synthesis inhibitor clindamycin triggered a stress response in *S. aureus* via alternative sigma factor B ( $\sigma^B$ ). This correlated with elevated biofilm production and was specific for the *S. aureus* USA300 clonal lineage. Wong and colleagues now show that a subinhibitory concentration of clindamycin under nonbiofilm conditions triggered a similar transcriptional response in CA-MRSA ST30 strains, resulting in the upregulation of  $\sigma^B$  as well as *agr* and the PSM $\alpha$  genes. In a next step, it will be interesting to see whether *S. aureus* ST30 strains also display clindamycin-dependent biofilm induction, as we observed for USA300 LAC and two USA300 clinical isolates derived from skin and soft tissues infections. The *VraSR* two-component system (TCS) is activated by cell wall-active antibiotics as well as cell wall hydrolysis or cell wall synthesis inhibition and regulates the cell wall stress response (3–6). Wong and colleagues showed that inactivation of the *VraSR* TCS modulated the transcription of several genes of *S. aureus* subjected to a subinhibitory concentration of clindamycin. We showed that clindamycin induced the upregulation of the murein hydrolase *AtlA* and triggered changes in cell morphology (2), which may lead to cell wall stress in *S. aureus*. Thus, subinhibitory concentrations of clindamycin might trigger a cell wall stress response via the *VraSR* TCS. The increased expression of *atlA* that we observed in the biofilm after clindamycin addition may be balanced by the induction of the *VraSR*-dependent cell wall stress response, resulting in downregulation of *atlA* and upregulation of cell wall biosynthesis enzymes, counteracting cell lysis. Increased biofilm formation has previously been connected to cell wall stress induced by cell wall-active antibiotics in various studies (7–9). Further investigations will be necessary to clarify the role of the *VraSR* TCS in the *S. aureus* response to subinhibitory concentrations of clindamycin and its role in biofilm induction.

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This is a response to a letter by Wong et al. (<https://doi.org/10.1128/AAC.02254-16>).

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